

Faculty / Presenter Disclosure

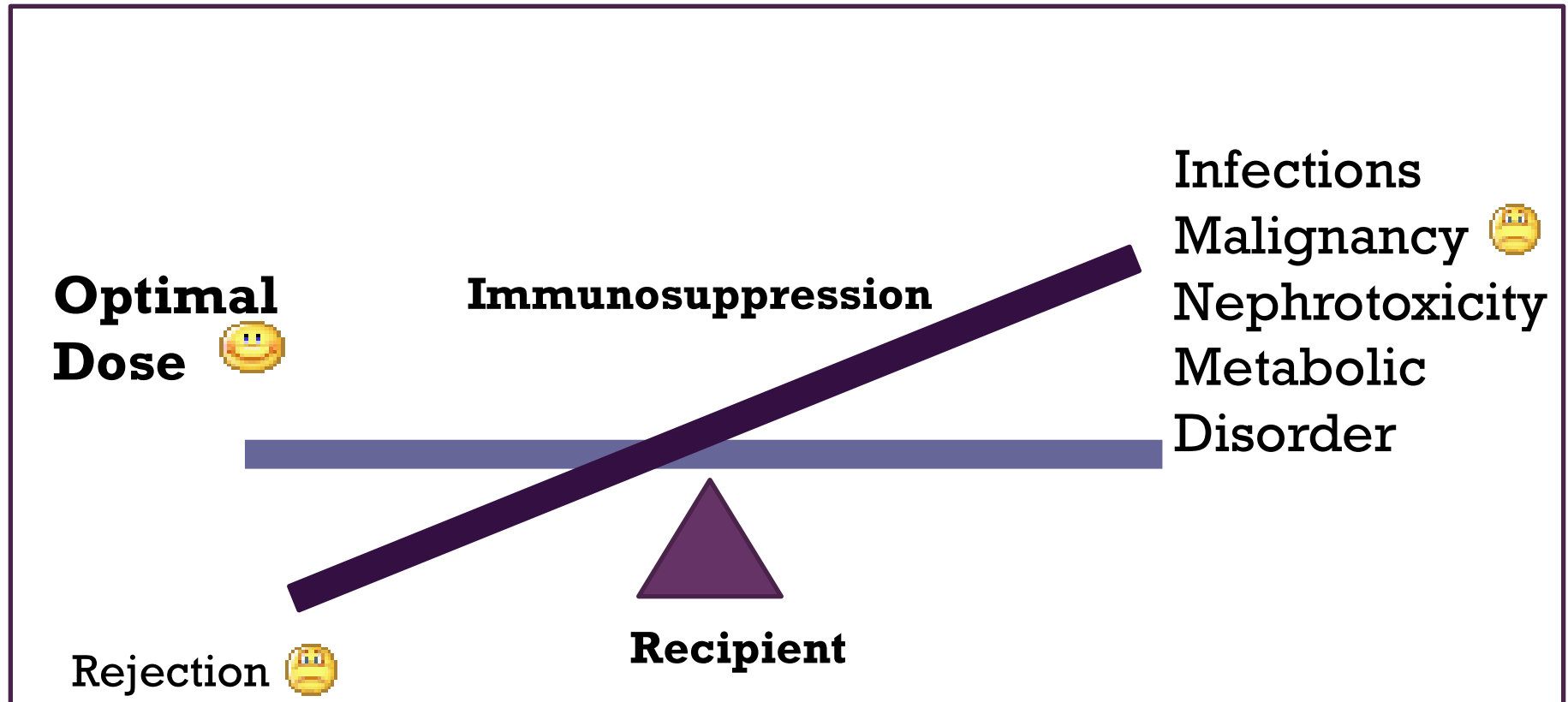
- Faculty: Maria Hernandez Fuentes
- Relationships with commercial interests:
 - Other: Employee of UCB Pharma
 - No off-label (or on-label) use of any product from UCB will be discussed in my presentation.

State of the art: Fingerprints of tolerance

Dr Maria Hernandez-Fuentes



The everlasting Challenge



How would finding tolerance help?

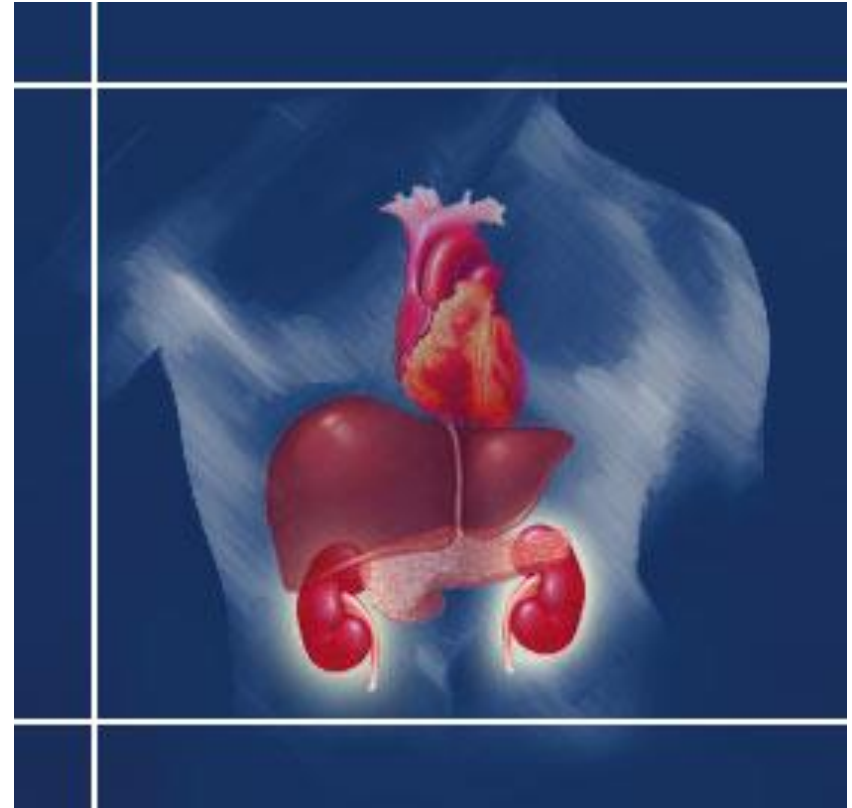


Holy Grial in clinical
Transplantation:

Tolerance

<http://www.ncbi.nlm.nih.gov/books/NBK26921/>

Donor-specific
unresponsiveness in the
context of otherwise normal
immune responses



Why Biomarkers of Tolerance?

- To identify *spontaneous* “tolerant-phenotype”
 - *Personalised medicine*: could those patients, identified as such, be optimally maintained with less immunosuppression?

- Evaluate novel “tolerance-inducing” therapies (cell therapy, regenerative therapies, etc)
 - would these patients display the same markers as patients with *spontaneous* tolerance?

- Better understand allo-immune regulation
 - would such markers allow us to understand better mechanisms of tolerance?

2002 – 2010 Indices of Tolerance & ITN

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Saskia Stevenson
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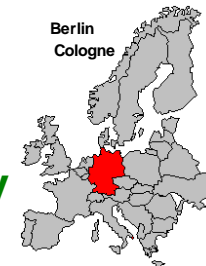
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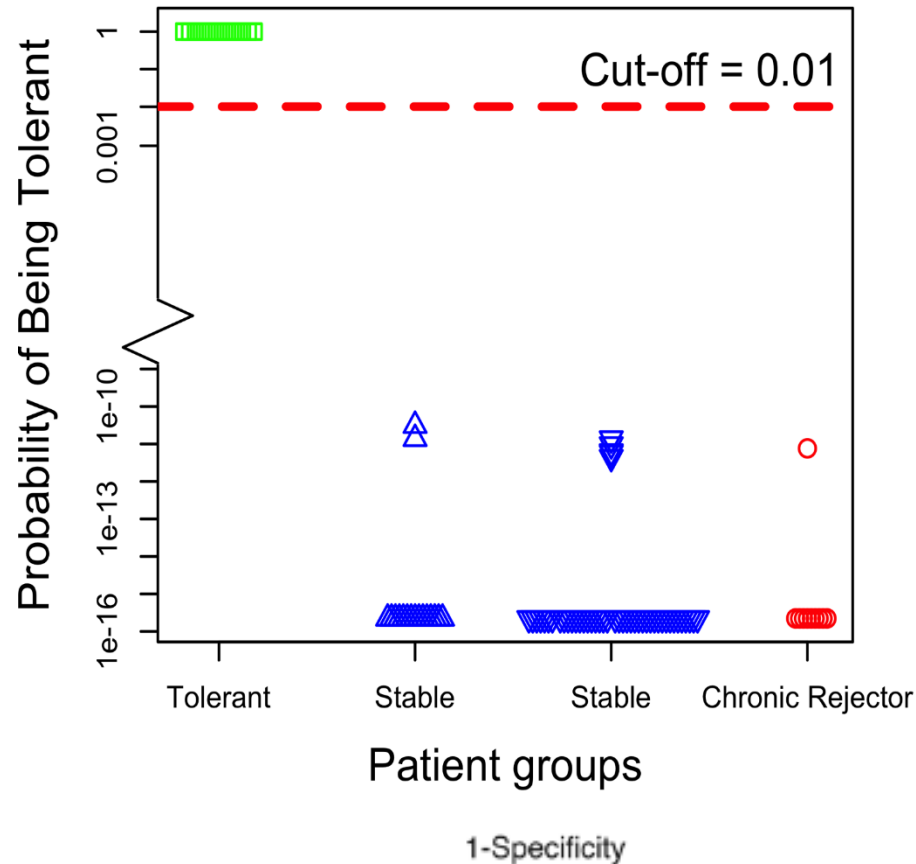
Immune Tolerance Network



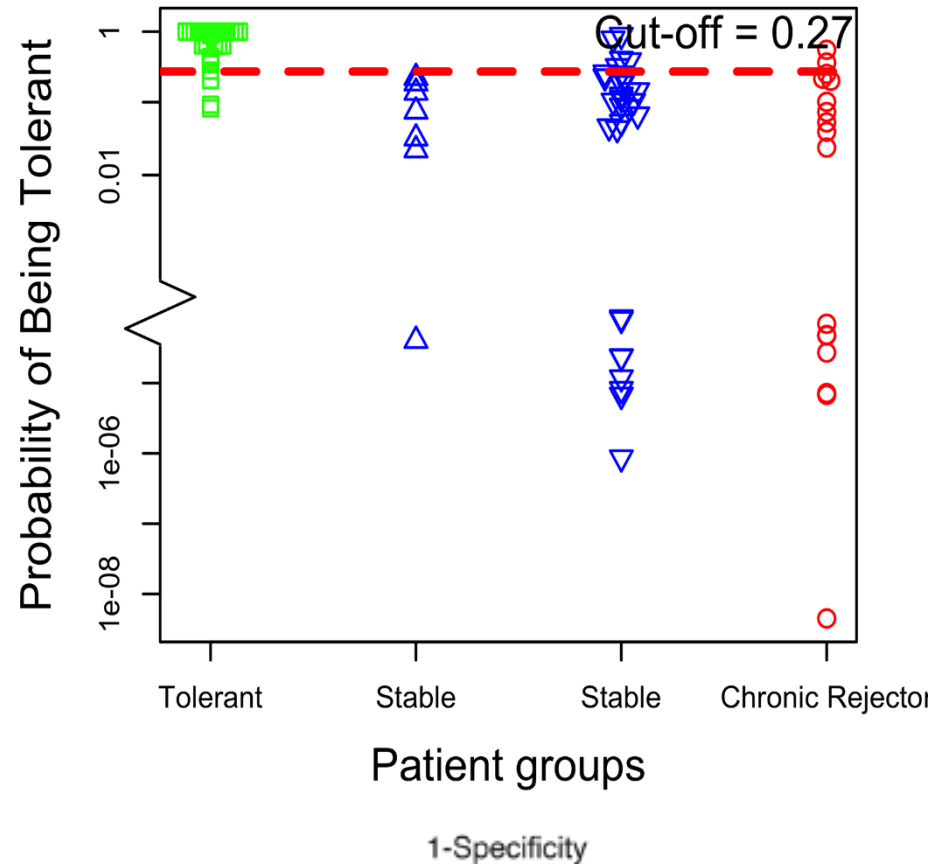
Vicki Seyfert
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Ignacio Sanz
Wei Chungwen
James Roger

Predicted Probability of being tolerant

EU Sample

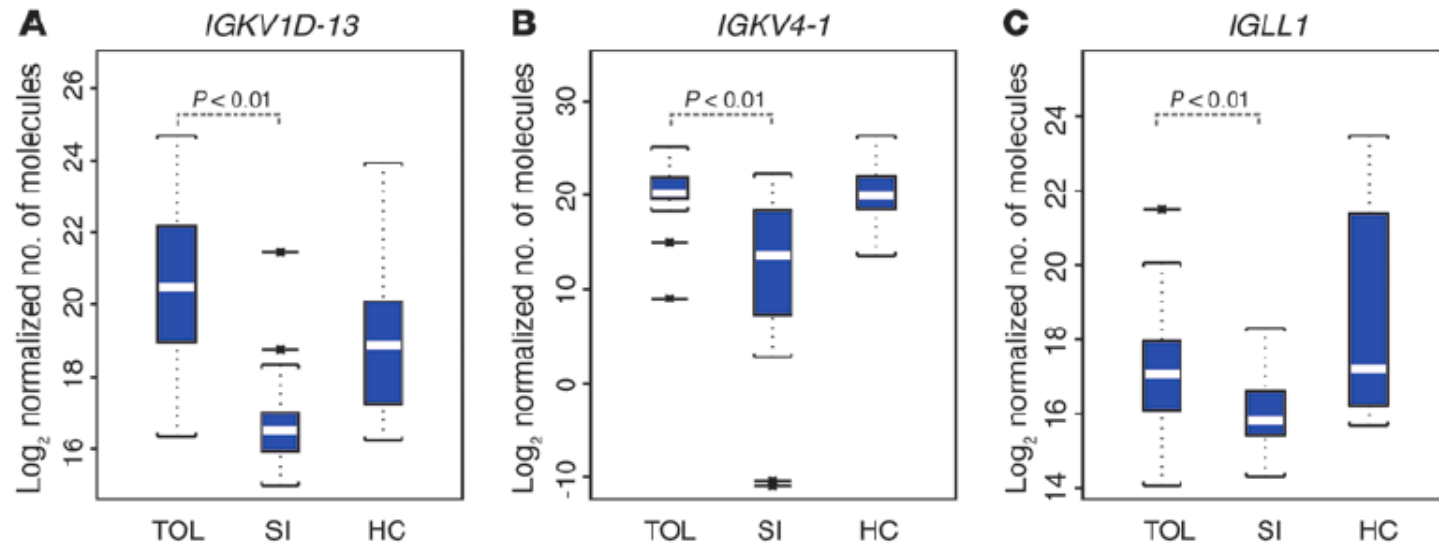


US Sample



Identification of a B cell signature associated with renal transplant tolerance in humans

Kenneth A. Newell,¹ Adam Asare,^{2,3} Allan D. Kirk,¹ Trang D. Gisler,^{2,3} Kasia Bourcier,^{2,3} Manikkam Suthanthiran,⁴ William J. Burlingham,⁵ William H. Marks,⁶ Ignacio Sanz,⁷ Robert I. Lechler,^{8,9} Maria P. Hernandez-Fuentes,^{8,9} Laurence A. Turka,^{3,10} and Vicki L. Seyfert-Margolis,^{3,11} for the Immune Tolerance Network ST507 Study Group



Tolerant subjects showed increased expression of multiple B cell differentiation genes.

A set of just 3 of these genes distinguished tolerant from non-tolerant recipients in a unique test set of samples.

Newell K, et al www.immunetolerance.org, et al. *J CLIN INVEST* (2010) 120 (6): 1836-47

Identification of a peripheral blood transcriptional biomarker panel associated with operational renal allograft tolerance

Sophie Brouard^a, Elaine Mansfield^{b,c}, Christophe Braud^a, Li Li^b, Magali Giral^a, Szu-chuan Hsieh^b, Dominique Baeten^{a,d}, Meixia Zhang^{b,e}, Joanna Ashton-Chess^a, Cécile Braudeau^a, Frank Hsieh^f, Alexandre Dupont^a, Annick Pallier^a, Anne Moreau^g, Stéphanie Louls^a, Catherine Ruiz^a, Oscar Salvatierra^b, Jean-Paul Soulillou^{a,j}, and Minnie Sarwal^{b,k}



Table 1. Demographic summary of patient groups (median and range)

	Training groups			Test groups					
	TOL	CR	N	TOL-test	CR-test	MIS	STA	AR	N-test
Number	5	11	8	12	11	10	12	14	8
Age, years	67 58–73	56 28–75	23 11–27	37.5 20–87	52 10–59	55.5 28–83	49 31–67	20 16–24	46 30–66
% Male	80	63.60	37.5	75	63.6	54.50	58	64.20	0
Time post-transplant, months									
Mean	178	59	NA	137	48	139.5	172	12	NA
Range	108–360	20–158		86–372	11–158	47–262	48–269	0.5–108	
Serum creatinine, $\mu\text{M/liter}$									
Mean	122	244	NA	115	244	98.5	107	152	NA
Range	82–139	127–492		70.4–149.6	100–686	64–161	63–147	110–704	

From microarray results a “tolerant footprint” of 49 genes.

These biomarkers were applied for prediction of operational tolerance by microarray and real-time PCR in independent test groups.

33 of 49 genes correctly segregated tolerance and chronic rejection phenotypes with 99% and 86% specificity.

The expression signature suggests that **TGF- might contribute to this process**, possibly by regulating specific phenotypes of peripheral regulatory T cells or altering the threshold for T cell activation

Proc Natl Acad Sci USA. **2007** 25;104(39):15448–53



Genetic Analysis & Monitoring of Biomarkers of Immunological Tolerance

GAMBIT study



The GAMBIT Consortium. NIHR CRN

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Biomarkers of Tolerance:

Patient groups in retrospective cohort;
aim to find the *prevalence of “tolerance”*

Tolerant recipients



Stable function
No immunosuppression

n=14

Stable recipients



Stable function

Little immunosuppression
Medium i-sup
High i-sup

n=190

Chronic Rejection



Graft Dysfunction

Medium i-sup
High i-sup

n=36

Index Group

Control Groups

Immunosuppression in GAMBIT

CNI	Azathioprine vs MMF	Prednisone	N	Percentage #
Cyclosporine	Azathioprine	No	25	13.2
Cyclosporine	Azathioprine	Yes	7	3.7
Cyclosporine	MMF	No	29	15.3
Cyclosporine	MMF	Yes	12	6.3
Cyclosporine	None	No	9	4.7
Cyclosporine	None	Yes	6	3.2
Tacrolimus	Azathioprine	No	11	5.8
Tacrolimus	Azathioprine	Yes	1	0.5
Tacrolimus	MMF	No	24	12.6
Tacrolimus	MMF	Yes	8	4.2
Tacrolimus	None	No	3	1.6
Tacrolimus	None	Yes	4	2.1
None	Azathioprine	No	1	0.5
None	Azathioprine	Yes	17	8.9
None	MMF	Yes	12	6.3
None	None	Yes	12	6.3
None	None	No	1 *	0.5

Percentage from all stable patients

* Patient on Sirolimus single therapy

Note: Information of drug regimen was missing for 8 of the stable patients (4.2%)

Retrospective
Study

17 therapy groups

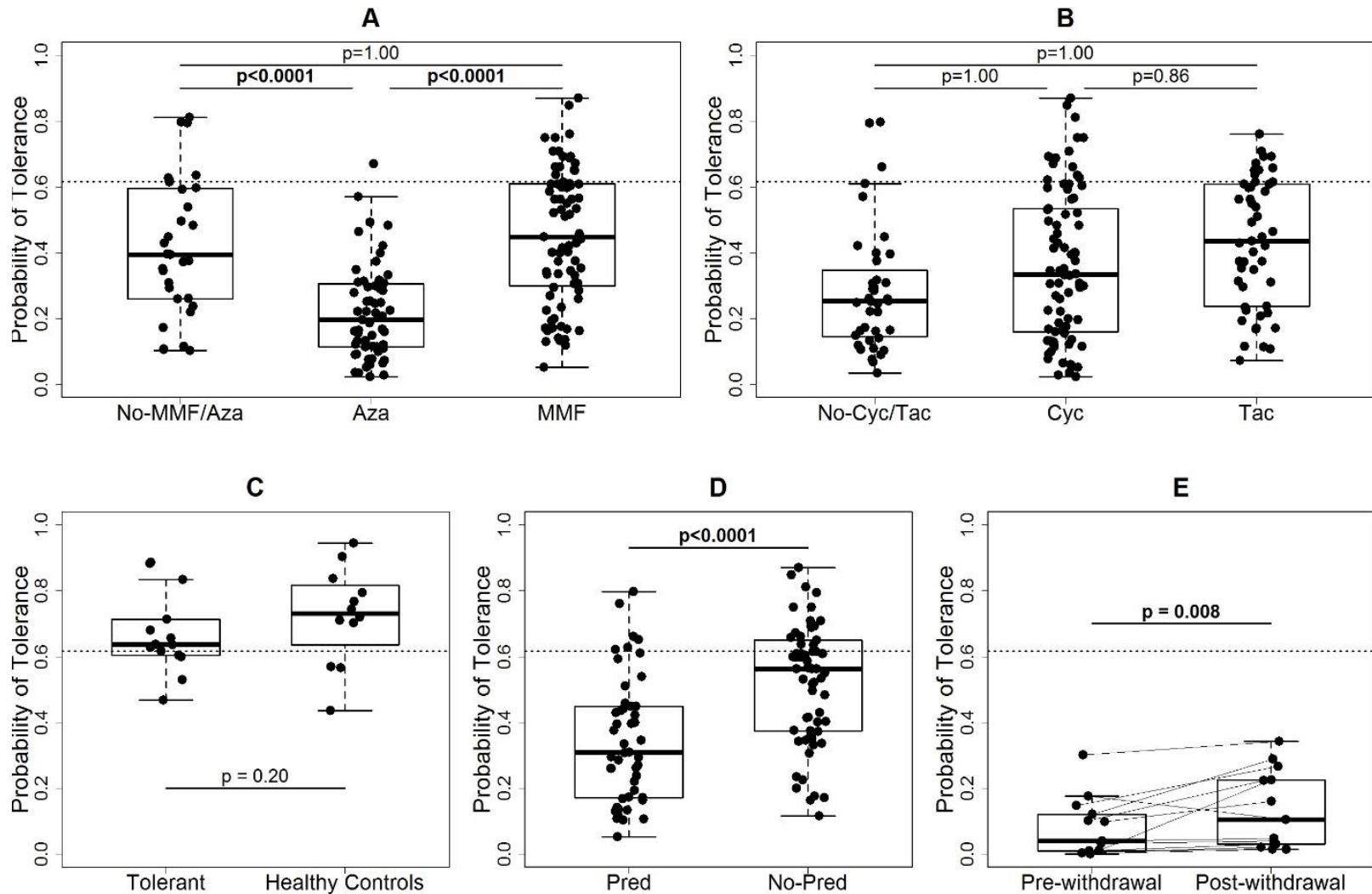
Unadjusted effect of immunosuppression in gene expression

	Pred	Cyc	Tac	Aza	MMF
PNOC	0.11	0.10	0.041	0.76	1.00
CD79b	2.1×10^{-04}	1.00	0.12	8.1×10^{-04}	0.94
TCL1A	1.9×10^{-06}	0.17	0.020	6.7×10^{-16}	1.00
H3ST1	1.3×10^{-04}	0.30	0.14	3.6×10^{-05}	0.20
SH2DB1	0.42	1.00	1.00	$< 2.0 \times 10^{-16}$	0.11
TLR5	4.0×10^{-03}	1.00	0.095	1.00	1.00
MS4A1	3.0×10^{-03}	0.73	0.18	1.1×10^{-04}	1.00
FCRL1	1.7×10^{-04}	1.00	0.73	1.1×10^{-10}	1.00
FCRL2	5.7×10^{-04}	1.00	0.15	1.6×10^{-05}	1.00
FoxP3/A Mann	0.69	0.16	9.0×10^{-03}	1.00	1.00

Rebollo-Mesa, et al AJTx 2016



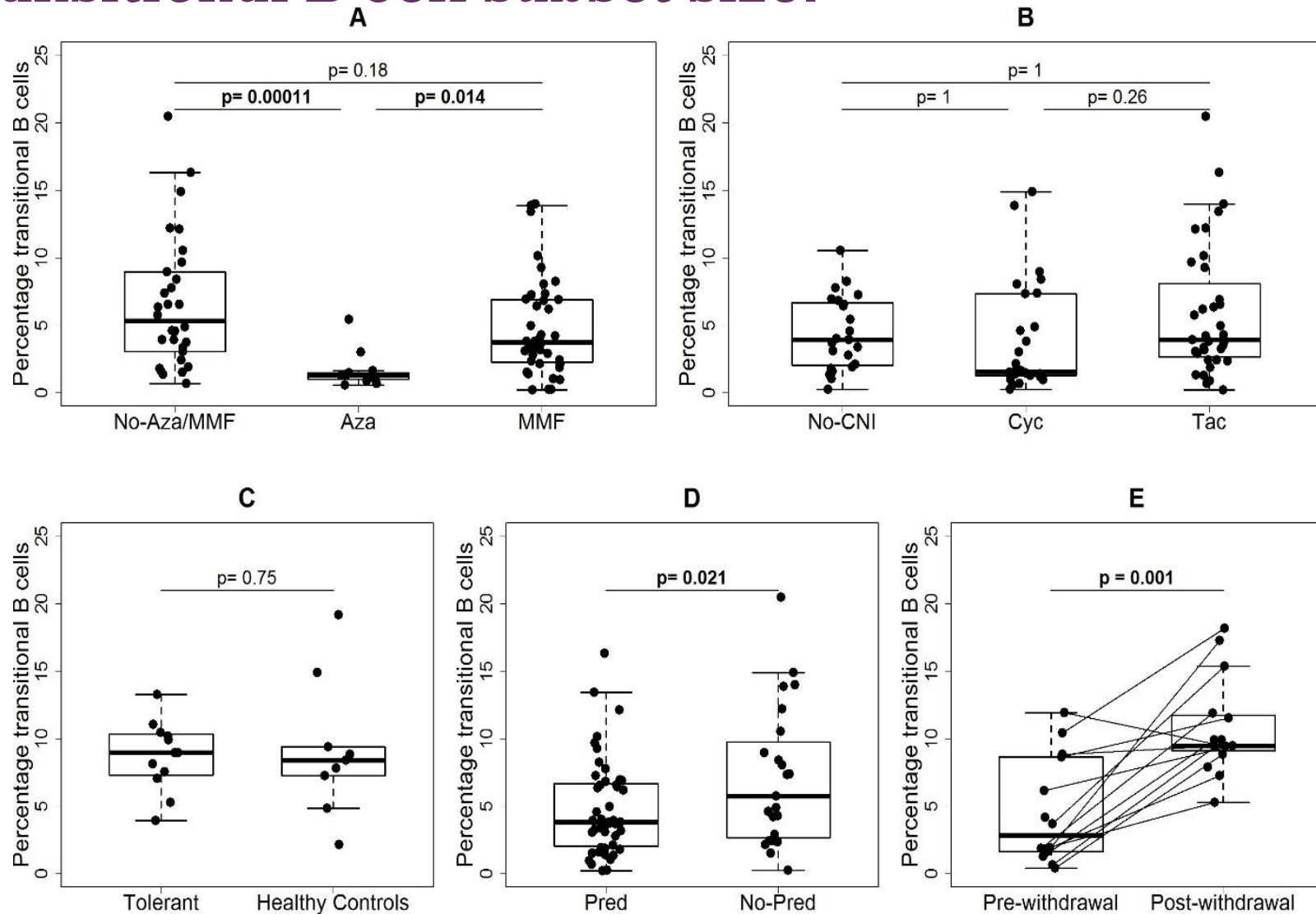
Immunosuppressants had a confounding effect on the expression of IoT gene set. RT-PCR



Rebollo-Mesa, et al AJTx 2016



Immunosuppressants affected the transitional B cell subset size.



Rebollo-Mesa, et al AJTx 2016

Conclusions 1

- Immunosuppression drugs the patients are taking (Aza + Pred)
 - have a major effect on the expression of the chosen genes
 - major effect on the size of the Transitional B cell compartment



dreamstime

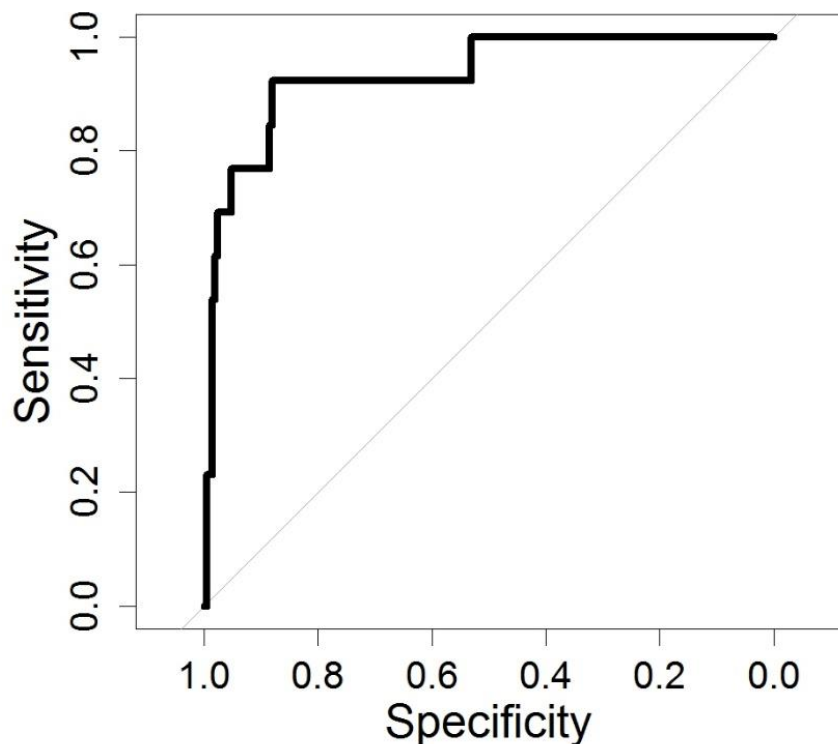
Rederived IS-independent gene-signature : IoT arrays

Validation using Fluidigm platform on GAMBIT samples.

EXPRES.	Molecular Function	Biological Processes	Documented protein expression in
↓ ATXN3	Ubiquitin-specific protease activity	Protein metabolism	Caudate Nucleus, Cerebellum Frontal Cortex, Pons, Ubiquitous
↓ BCL2A1	Receptor signaling complex scaffold activity	Apoptosis	B cell, Bone Marrow, Colon, Intestine, Leucocyte, Lymph node, Ovary, Spleen, T cell
↓ EFF1A1	Transcription regulator activity	Regulation of cell cycle	B cell, Islets of Langerhans, Lacrimal gland, Leukocyte, Monocyte, Neutrophil, Plasma, Saliva, Semen, Skeletal muscle, Tear
↓ TNFAIP3	Transcription regulator activity	Regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolism	Macrophages
↓ NFKBIA	Transcription regulator activity	Regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolism	Neutrophil, T cell
↑ GEMN7	Ribonucleoprotein	Regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolism	Spinal cord tissues
↑ IGLC1	Antigen binding	Immune response	B lymphocytes
↑ MS4A4	Unknown	Unknown	Intestine and colon
↑ RAB40	GTPase activity	Cell communication Signal transduction	Platelets, Liver, Heart, Kidney, Plasma

Rebollo-Mesa, et al AJTx 2016

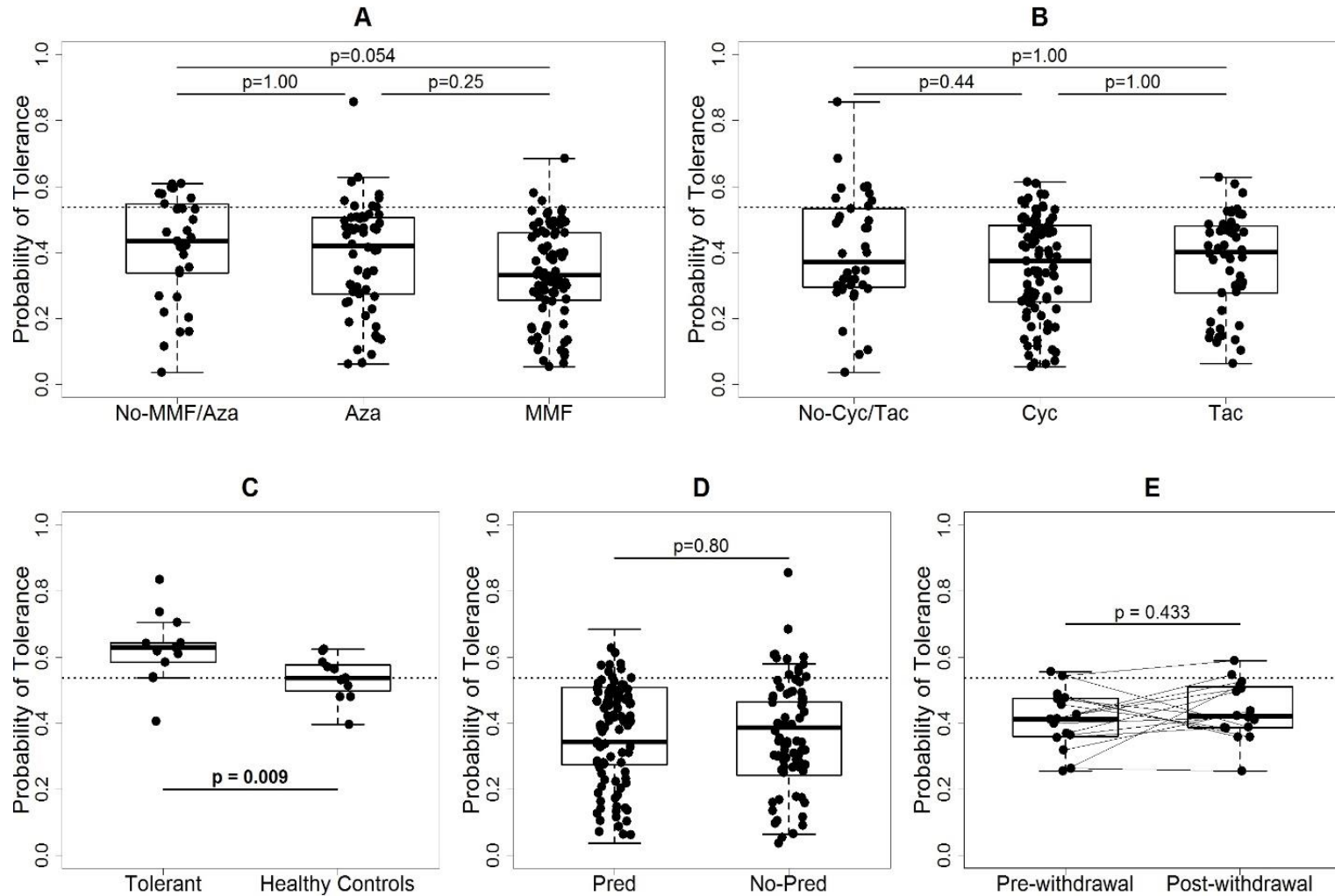
Diagnostic characteristics of IS-independent signature of tolerance



Performance measure	IS-free signature
AUC	0.93
95% CI of AUC	0.86-1.00
CV AUC	0.81
Cutoff	0.54
Sensitivity	0.92
Specificity	0.88
AUC Timepoint 2	0.83
95% CI of AUC Timepoint 2	0.67 - 0.99

Rebollo-Mesa, AJTx et 2016

Robust immunosuppression independence of IS-independent set



Rebollo-Mesa, et al AJTx 2016

Conclusions 3

- P(tolerance) unchanged after steroid withdrawal: New-signature highlights **natural counter-regulatory mechanisms**, and excludes the alterations of the immune effector pathways transiently activated or inhibited by IS drugs.
- This signature is “**tolerance –specific**” as it is significantly different from Healthy Controls
- The use of **robust statistical methods** that prevent false-positive results, and control confounding is essential prior to translation of clinical prediction models.

see commentary on page 875

A common gene signature across multiple studies relate biomarkers and functional regulation in tolerance to renal allograft

Daniel Baron^{1,2,3}, Gérard Ramstein⁴, Mélanie Chesneau^{1,2,3}, Yann Echasserieau^{1,2,3}, Annaick Pallier^{1,2,3}, Chloé Paul^{1,2,3}, Nicolas Degauque^{1,2,3}, Maria P. Hernandez-Fuentes⁵, Alberto Sanchez-Fueyo⁶, Kenneth A. Newell⁷, Magali Giral^{1,2,3}, Jean-Paul Soulillou^{1,2,3}, Rémi Houlgatte^{8,9,10} and Sophie Brouard^{1,2,3,10}

CLINICAL RESEARCH

www.jasn.org

A Three-Gene Assay for Monitoring Immune Quiescence in Kidney Transplantation

Silke Roedder,* Li Li,[†] Michael N. Alonso,[‡] Szu-Chuan Hsieh,* Minh Thien Vu,* Hong Dai,* Tara K. Sigdel,* Ian Bostock,[§] Camila Macedo,^{||} Diana Metes,^{||} Adrianna Zeevi,^{||} Ron Shapiro,^{||} Oscar Salvatierra,[‡] John Scandling,[‡] Josefina Alberu,[§] Edgar Engleman,[‡] and Minnie M. Sarwal*

The questions

- Are any of these found in tolerance inducing strategies?

- Newell KA, et al AJTx 2015; 15: 2908–2920 .

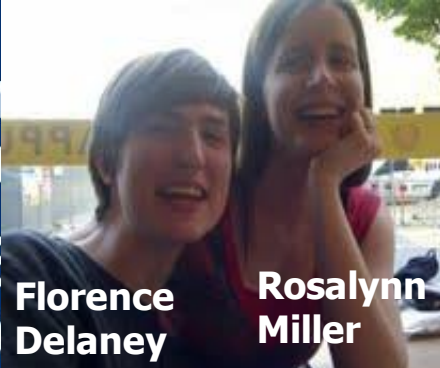
- Have we learned novel mechanisms of tolerance?

.... Transitional B cells / Role of steroid pathway

- Increased CD40 Ligation and Reduced BCR Signalling Leads to Higher IL-10 Production in B Cells From Tolerant Kidney Transplant Patients. Nova-Lamperti E, et al Transplantation. 2017 Mar;101(3):541-547
 - IL-10-produced by human transitional B-cells down-regulates CD86 expression on B-cells leading to inhibition of CD4+T-cell responses. Nova-Lamperti E, et al . Sci Rep. 2016 Jan 22;6:20044.

- Are any of these “true” biomarkers of tolerance?.

- Clinical trials of weaning are needed = controversial



Guy's and St Thomas' NHS
Foundation Trust and King's
College London's
**Comprehensive Biomedical
Research Centre**



Pioneering better health for all



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Nicola Smallcombe
Jennifer Mullan
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Tissue typing at Guy's Hospital:

Dr Robert Vaughn, Livvy Shaw & Team